Phylogeographic relationships between the mosses *Exsertotheca intermedia* from Macaronesian islands and *Neckera baetica* from southern glacial refugia of the Iberian Peninsula

Isabel Draper^{1,*}, Juana M. González-Mancebo², Olaf Werner³, Jairo Patiño^{2,4} & Rosa M. Ros³

- ¹⁾ Departamento de Biología, Facultad de Ciencias, Universidad Autónoma de Madrid, c/ Darwin, 2, ES-28049 Madrid, Spain (*corresponding author's e-mail: isabel.draper@uam.es)
- ²⁾ Departamento de Biología Vegetal, Facultad de Biología, Universidad de La Laguna, ES-38271 Tenerife, Spain
- ³⁾ Departamento de Biología Vegetal, Facultad de Biología, Universidad de Murcia, Campus de Espinardo, ES-30100 Murcia, Spain
- ⁴⁾ Present adress: Dep. Environmental Sciences and Management, Institute of Botany, University of Liège, B22 Sart Tilman, BE-4000 Liège, Belgium

Received 9 Feb. 2011, revised version received 17 Mar. 2011, accepted 18 Mar. 2011

Draper, I., González-Mancebo, J. M., Werner, O., Patiño, J. & Ros, R. M. 2011: Phylogeographic relationships between the mosses *Exsertotheca intermedia* from Macaronesian islands and *Neckera baetica* from southern glacial refugia of the Iberian Peninsula. — *Ann. Bot. Fennici* 48: 133–141.

The phylogeographic relationships between the Macaronesian islands and southern glacial refugia of the Iberian Peninsula was studied using molecular markers and two mosses as model species: *Exsertotheca intermedia* from Macaronesia and the recently described *Neckera baetica* from the Iberian Peninsula. The new combination *Exsertotheca baetica* (Guerra) Draper, González-Mancebo, O. Werner, J. Patiño & Ros is proposed for the latter. The genus *Neckera* has recently undergone considerable systematic and taxonomic changes, but our results show that the European species *N. cephalonica* and *N. pumila* belong in *Neckera s. stricto*.

Introduction

The Macaronesian archipelagoes (Azores, Canary Islands and Madeira) are traditionally regarded as outstanding refugia for relict Tertiary lineages (Engler 1879, Axelrod 1975, Bramwell 1976, Sunding 1979, Rodríguez-Sánchez & Arroyo 2008, Rodríguez-Sánchez *et al.* 2009). However, mesic areas within the Mediterranean basin have also enabled long-term survival of some species in several glacial refugia (Vogel *et al.* 1999, Thompson 2005, Mejías *et al.* 2007, Rodríguez-Sánchez *et al.* 2008, 2009), as have also some Atlantic areas in the Iberian Peninsula (Sunding 1979, Désamoré *et al.* 2011).

The south-west of the Iberian Peninsula constitutes one of the main glacial refugia for the European fauna and flora (Hewitt 2000).

The Macaronesian region is characterized by high rates of endemicity in a vast array of organisms (Juan *et al.* 2000). This also applies to bryophytes, although this group generally exhib-

its much lower levels of endemism than vascular plants (Sotiaux et al. 2009, Aigoin et al. 2009, Vanderpoorten et al. 2011). The presence of endemics is precisely at the basis of the biogeographical concept of Macaronesia (Engler 1879, Sunding 1979). However, in addition to the strict Macaronesian endemic taxa, there is a group of species present both on the Macaronesian islands and in some glacial refugia of the Iberian Peninsula. Such taxa are called Ibero-Macaronesian endemics. In comparison with the strict Macaronesian endemics, there are few plant species with this type of distribution, such as Convolvulus fernandesii among the vascular plants (Carine et al. 2004), and the bryophytes Frullania azorica, Porella canariensis, Tetrastichium fontanum, T. virens, and Isothecium algarvicum. Until very recently, also Exsertotheca intermedia (Olsson et al. 2011) (= Neckera intermedia: see González-Mancebo et al. 2008) was considered an Ibero-Macaronesian endemic. However, the study of Guerra et al. (2010) rejected this concept, and they described a new species from the Iberian Peninsula, Neckera baetica, which had previously been overlooked.

Some previous studies on the Neckeraceae (Tsubota et al. 2002, Ignatov et al. 2007, Olsson et al. 2011) revealed that the genus Neckera as traditionally understood is paraphyletic. The results obtained by Olsson et al. (2011) suggest that it is phylogenetically divided into four clades that should be recognized as separate genera. The clade containing the generitype N. pennata is called Neckera s. stricto. Some species previously placed in Neckera became grouped in Forsstroemia. Finally, the rest of the 'Neckera' species studied by Olsson et al. (2011) fall in two well-supported clades, and so those authors established two new genera to accommodate them: Alleniella (with ten species, including the European A. besseri and A. complanata) and Exsertotheca (including two species, E. crispa and E. intermedia).

In order to analyse the phylogeographic relationships between the Macaronesian islands and southern glacial refugia of the Iberian Peninsula we selected the Macaronesian endemic *Exsertotheca intermedia*, and the recently described *Neckera baetica* from the Iberian Peninsula. The first species was considered a suitable case study due to its relative abundance in Macaronesia (Hedenäs 1992). It has been reported from all of the Macaronesian archipelagoes (Hedenäs 1992, Patiño Llorente & González-Mancebo 2005, Frey et al. 2006) and was until very recently also thought to occur in the Iberian Peninsula (Guerra et al. 1980, 2003, Düll 1992, Frey et al. 2006). Exsertotheca intermedia was described as Neckera intermedia from material collected on Tenerife (Canary Islands), on the basis of the transversely undulate, oblong stem leaves with a rounded to obtuse apex (Bridel 1812). Due to its morphological variability several other taxa were described, but were later synonymised by Van der Wijk et al. (1964), De Sloover (1977) and Hedenäs (1992): Neckera bolleana, N. elegans, N. elegans var. laevifolia and N. laeviuscula. The recent segregation of the Iberian specimens as a separate species N. baetica (Guerra et al. 2010) renders E. intermedia a Macaronesian endemic, and arouses questions on the origin of the recently described species.

The description of *N. baetica* coincides temporally with our research on the phylogeographic relationships between the Macaronesian islands and the southern glacial refugia in the Iberian Peninsula using *Exsertotheca* species. Guerra *et al.* (2010) used nuclear ITS sequences and provided a morphological description of the species, but in our opinion some of the characters they propose are not completely useful for distinguishing the species. Therefore, the delimitation of *N. baetica* should be clarified under the framework of a most appropriate molecular approach (*see* Olsson *et al.* 2011).

Our main aim is to analyse the phylogenetic relationship between populations of E. *interme*dia from the Macaronesian islands and those of N. baetica from the Iberian Peninsula to infer the evolutionary origin of the latter. Secondly, we aim to clarify the present generic concept of Neckera s. lato species in Europe in the light of the proposal by Olsson et al. (2011).

Material and methods

Molecular study

Samples from seventeen populations of E. inter-

media and five populations of N. baetica were included in the molecular study. In addition, we included samples of other species traditionally included in Neckera and occurring in the Iberian Peninsula: Alleniella besseri (= Neckera besseri, two populations), A. complanata (= N. compla*nata*, six populations), *Exsert otheca crispa* (= N. crispa, eight populations), Neckera cephalonica (two populations), and N. pumila (four populations). Other two Neckera s. stricto species that are present in the Iberian Peninsula (Sérgio et al. 2006), N. menziesii and N. pennata, are both phylogenetically clearly separated from E. intermedia (Olsson et al. 2011). Molecular sequences of these two species that are available from Gen-Bank were also included in the present study in order to picture the phylogenetic relationship of all the Iberian species. Finally, a sample from one population of Leptodon smithii was used as outgroup. Details for all the sequenced specimens are provided in the Appendix.

We used two molecular markers, the nuclear ITS and the chloroplast intron trnL-F. Total DNA was extracted using the NaOH method of Werner et al. (2002). PCR was performed in an Eppendorf Mastercycler using PuReTaqTM Ready-To-Go[™] PCR Beads (GE Healthcare) in a 25 μ l reaction volume according to the manufacturers' instructions. For the two molecular markers, the PCR programs given below were initiated by a melting step of 5 min at 95 °C and were followed by a final extension period of 8 min at 72 °C. For the internal transcribed spacer (ITS) the PCR program employed was 35 cycles of 30 s at 95 °C, 30 s at 52 °C and 1 min 45 s at 72 °C, with the primers 'AB101F' and 'AB102R' (Douzery et al. 1999) or '25R' (Stech & Frahm 1999) and '18F-ISO'. The latter is slightly modified from the '18F' designed by Stech and Frahm (1999) and is constituted of the following base sequence: 5'GGA TGG AGA AGT CGT AAC AAG G 3'. For the chloroplast intron trnL-F, 35 cycles of 30 s at 95 °C, 45 s at 51 °C and 1 min 30 s at 72 °C were employed, with the primers 'TabC' and 'TabF' (Taberlet et al. 1991). Aliquots (5 μ l) of the amplification products were visualized on 1% agarose gel. Successful amplifications were cleaned with a GenElute™ PCR Clean-Up kit (Sigma-Aldrich Biotechnology). The amplification primers were

used in the sequencing reactions with the Big Dye sequencing kit and separated on an ABI-Prism 3700 using standard protocols.

Nucleotide sequence fragments were edited and assembled for each DNA region using PhyDE v0.995 (Müller *et al.* 2006). The assembled sequences were manually aligned, on the basis of the criteria provided by Kelchner (2000), and regions of incomplete data at the beginning and end of the sequences were excluded from subsequent analyses. The sequence alignments used in the analyses are available on request.

Phylogenetic analyses

Phylogenetic reconstructions were made on the basis of Maximum Parsimony (MP) and Bayesian Inference (BI) optimality criteria using the programs TNT 1.0 (Goloboff et al. 2003) for MP, and MrBayes 3.1 (Huelsenbeck & Ronquist 2001, Ronquist & Huelsenbeck 2003) for BI. MP phylogeny was analysed by a traditional heuristic search, selecting tree bisection reconnection (TBR) as swapping algorithm, with one tree held in memory. All characters were equally weighted. Clade support was assessed via nonparametric bootstrapping using the default settings in TNT, except for the number of replicates, which was set to 1000. The output results requested were the absolute frequencies. For all generated maximum parsimony trees the consistency index (Ci) and retention index (Ri), as well as tree length were calculated.

The nucleotide substitution model used for the BI analysis was selected with jModeltest 0.1.1 (Posada 2008), based on Bayesian information criteria. BI analysis used one cold and three incrementally heated Monte Carlo Markov chains (MCMC) on two simultaneous runs. MCMC run until the standard deviation of split frequencies was below 0.01 (1 000 000 generations), with one tree sampled every 100th generation, each using a random tree as a starting point and a temperature parameter value of 0.2 (the default in MrBayes). The first 25% of the total sampled trees of each run were discarded as burn-in, in order to achieve the MCMC log-likelihoods that had become stationary and converged. The remaining sampled trees were used to construct a Bayesian consensus tree and to infer posterior probabilities (PP).

While gaps in non-coding regions are sometimes difficult to assess (Kelchner 2000), all the analyses were run both with the insertions and deletions coded as informative characters and without coding the insertions and deletions. The indels were coded using Simple Indel Coding (SIC) strategy, by Simmons and Ochoterena (2000), as implemented in SeqState (Müller 2004).

Results

Molecular analyses

The nucleotide substitution model that best suits our data set was K80+G. Length variation for the ITS dataset (ITS1, 5.8S rDNA, ITS2) ranges from a low of 725 nucleotides to a high of 777 nucleotides (missing data, length polymorphic regions, 3' end of SSU rDNA and 5' end of LSU rDNA excluded), of which 64 sites are variable, and of these 45 are parsimony-informative. Out of the 462 to 463 nucleotides included in the trnL-F, 19 are variable and 16 of these are parsimony informative. The simple indel coding doubled the number of parsimony-informative characters in the case of the nuclear data sets, increasing them with 34 characters. In the case of the chloroplast data sets, the simple indel coding yielded only one additional parsimonyinformative character.

The two optimality criteria used (MP and BI) yielded similar results in all the analyses run, although BI resolves some of the relationships that remain unclear under the MP analysis. Separate analyses of the nuclear and the chloroplast datasets result in clearly congruent tree topologies that differ only in unsupported clades, and so the ITS and trnL-F datasets are combined. The topology of the trees resulting from the analyses coding and without coding the insertions and deletions as informative characters are also congruent, although MP trees obtained without the indels coded are markedly shorter than those obtained with indels coded. The tree shown (Fig. 1) is therefore the result of the BI analysis of the dataset that combines the two molecular markers, not including the insertions

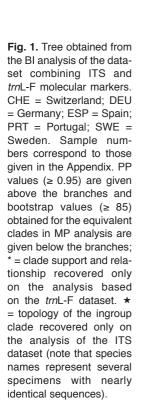
and deletions coded as informative characters. The MP analysis of the same combined dataset yields 693 equally parsimonious trees of 160 steps (Ci = 0.869, Ri = 0.954) and has an overall similar topology, albeit lacking significant support for some of the clades (bootstrap values for well supported clades are indicated in Fig. 1).

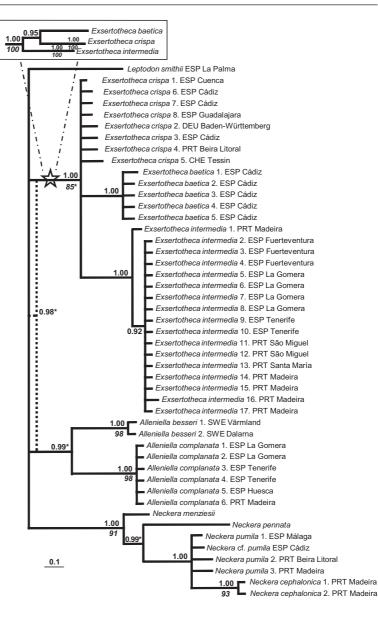
In all the analyses, the studied samples of N. baetica were positioned together in a clade that is clearly separated (PP = 1.00) from the clade in which all the E. intermedia samples from Macaronesia gather (Fig. 1). Moreover, when the ITS marker is separately analysed, E. crispa is resolved in a well supported clade (PP = 1.00, bootstrap value = 100), and the *N*. baetica populations are placed as a sister clade (with PP = 0.95) to this E. crispa group, instead of together with the Macaronesian E. intermedia, as was also observed by Guerra et al. (2010). Within the studied taxa, N. baetica is phylogenetically closely related to the Macaronesian E. intermedia and the European-Asiatic E. crispa. These three species constitute a well-supported clade in all the analyses (PP = 1.00), which according to the chloroplast marker trnL-F is sister to A. besseri and A. complanata. Finally, Neckera pumila and N. cephalonica are closely related to N. pen*nata* (PP = 0.99 according to the *trn*L-F marker) and N. menziesii (PP = 1.00, bootstrap value = 91), constituting a monophyletic basal clade, which is apparently not more closely related to Alleniella or Exsertotheca than the outgroup taxon Leptodon smithii.

Discussion

Our results (Fig. 1) fully agree with the generic circumscription proposed by Olsson *et al.* (2011). In addition, the study of two molecular markers, including one nuclear and one chloroplast, and the increased number of sequences analysed, give support to the description of a separate species for the Iberian Peninsula, as Guerra *et al.* (2010) suggested on the basis of the study of ITS markers of two specimens for each species included in their study. All the studied samples of *Exsertotheca crispa* and *E. intermedia* are grouped in a well-resolved clade, together with the samples of *Neckera baetica.*

0.1





The included specimens of Alleniella besseri and A. complanata constitute a separate clade, which is sister to the one including Exsertotheca. Finally, our samples of Neckera pumila and N. cephalonica fall into a clade containing Neckera s. stricto species (N. menziesii and N. pennata), and which is clearly separated both from Exsertotheca and from Alleniella.

The close relationship of the studied samples of *N*. *baetica* with *E*. *intermedia* and *E*. *crispa* justifies placing the recently described *N*. *baetica* into the genus *Exsertotheca*. In order to

better understand the phylogenetic position of *N. baetica*, we have also run MP and BI analyses of the ITS dataset used in the present study including the wider Neckeraceae ITS sampling of Olsson *et al.* (2011), available in GenBank. As expected, *N. baetica* falls into the *Exsertotheca* clade of Olsson *et al.* (2011), as sister clade of *E. crispa* and closely related to *E. intermedia*. The resulting trees show identical topologies to those shown in the present study and by Olsson *et al.* (2011) and are therefore not shown. Regarding the other samples sequenced for this study, our

Alleniella besseri and A. complanata specimens are grouped together with Olsson's et al. (2011) samples of the corresponding species. As for Neckera pumila and N. cephalonica, our samples fall into Olsson's et al. (2011) Neckera s. stricto clade, which precludes any taxonomic change for these two species not previously studied by Olsson et al. (2011). The interest of the confirmation of these two species as belonging to Neckera has to be highlighted in the taxonomic frame of the Neckeraceae shown by Olsson et al. (2011).

After our contribution, from the nine species of *Neckera* accepted by Hill *et al.* (2006) for Europe and Macaronesia, *N. oligocarpa* (known from Scandinavia, Austria and Italy according to Frey *et al.* 2006; also from North America according to Anderson *et al.* 1990) is the only one for which the generic placement remains to be tested by molecular data. Taxonomically, this species appears to be close to *N. pennata*, with which it has been synonymized (Ignatov & Afonina 1992) or treated at varietal rank (*N. pennata* var. *tenera*). We therefore think that it probably belongs to *Neckera s. stricto*, although the verification is still necessary.

Exsertotheca baetica (Guerra) Draper, González-Mancebo, O. Werner, J. Patiño & Ros, *comb. nova*

Neckera baetica Guerra, Nova Hedwigia 91: 259, f. 4-6, 7, 9, 12, 14. 2010.

We agree with Guerra et al. (2010) that Exsertotheca baetica is well separated from E. intermedia, not only by the studied molecular markers, but also morphologically. These authors consider that the species is mainly characterized by its large shoots (up to 35 cm), yellowish green, ovate-oblong leaves, which are transversely undulate or plane when dry, obtuse to rounded leaf apex, plane leaf margins, eporose upper and middle laminal cells with 1.5–2 μ m thick walls, and a slightly papillose and spiculose endostome. They characterize E. intermedia by its dark green-brown to coppery colour, mostly oblong or elongate oblong leaves, which are occasionally lightly constricted in the upper third and strongly transversely undulate,

porose laminal cells with the mid-leaf cells walls (2)3.5-3.7(4.5) µm thick, and strongly papillose, not spiculose endostome processes. Guerra et al. (2010) also state that E. baetica is similar to E. intermedia in the leaf apex, the plane leaf margins, and the size and shape of the laminal cells. Our morphological study allows us to add some morphological differences between these species. In E. baetica the plants are small to large (no more than 30-35 cm long), but never as large as E. intermedia, which can reach a length of 1 m in the Canary Islands. The stems of *E. baetica* are up to 3 mm wide including the leaves, whereas they usually reach 4-5 mm in E. intermedia. It has to be mentioned that both species are highly variable in size depending on the environmental conditions in Quercus canariensis forests (E. baetica) or in laurel forests (E. intermedia). The stem leaves are generally asymmetric, spirally arranged on the branches and only loosely complanate on the old stems in E. baetica, and slightly asymmetric and complanate in E. intermedia; the stem leaf apex is slightly denticulate in E. baetica and generally entire in E. intermedia. It is also important to mention that, according to our observations, E. baetica has a slightly to strongly concave leaf lamina, not undulate when moist, not undulate or rugose and with 1-2 undulations when dry, while the leaves of E. intermedia are slightly transversely undulate when moist and strongly so when dry. Therefore, the placement of E. baetica in the group of undulate-leaved Neckera species by Guerra et al. (2010) cannot be sustained.

The recent description of E. baetica, an Iberian endemic, renders E. intermedia a Macaronesian endemic. Morphologically E. baetica is closer to E. intermedia than to E. crispa, in spite of the fact that the sampled E. baetica populations form a sister clade to the sampled E. crispa populations (Fig. 1). Even if further studies are necessary in order to reliably assess the origin for E. baetica, this morphological similarity suggests that it shared a common ancestor with *E. intermedia*. It is possible that *E*. baetica represents a recently diverging species in a Mediterranean glacial refuge, as was recently proposed for the Corsican endemic Leptodon corsicus (Sotiaux et al. 2009). If so, the origin of E. baetica might be more associated with

ecological differentiation than with geographical disjunction, since the area where it thrives in the south of the Iberian Peninsula has a very different climate than the surrounding areas. The climate is Mediterranean, but with a strong oceanic influence, and it has already been pinpointed as one of the main hotspots for plant biodiversity, both because of the presence of a relict flora and for its high rate of endemism (Rodríguez-Sánchez et al. 2008). Nevertheless, another possible explanation for the presence of the species in this area is that it is a relict of an originally Tertiary species. Studies on vascular plants have shown that endemics in the southwest of the Iberian Peninsula have followed different evolutionary patterns, including both long-term isolation and multiple active processes of speciation (Rodríguez-Sanchez et al. 2008, 2009). Further studies are therefore necessary in order to be able to date the time of divergence of *E. baetica*.

Acknowledgements

We are grateful to M. Brugués (BCB), R. Gabriel (AZU), L. Hedenäs (S), D. Long (E) and A.J. Shaw (DUKE), for the loans that made possible the present study. We also want to thank S. Olsson, S. Huttunen, and J. Enroth for their useful comments and suggestions regarding the manuscript, and V. Mazimpaka and F. Lara for their assistance. This work has been carried out with Financial support from the Spanish Ministerio de Educación y Ciencia (CGL2008-00275/BOS and Juan de la Cierva program), the Ministerio de Medio Ambiente (129/2006), and the Canary Islands Government (P1042004-028).

References

- Aigoin, D., Devos, N., Huttunen, S., Ignatov, M. S., González-Mancebo, J. M. & Vanderpoorten, A. 2009: And if Engler was not completely wrong? Evidence for multiple evolutionary origins in the moss flora of Macaronesia. — *Evolution* 63: 3248–3257.
- Anderson, L. E., Crum, H. A. & Buck, W. R. 1990: List of mosses of North America North of Mexico. — *The Bryologist* 93: 448–499.
- Axelrod, D. I. 1975: Evolution and biogeography of Madrean-Tethyan sclerophyll vegetation. — Annals of the Missouri Botanical Garden 62: 280–334.
- Bramwell, D. 1976: The endemic flora of the Canary Islands: distribution, relationships and phytogeography. — In: Kungel, G. (ed.), *Biogeography and ecology in the Canary Islands*: 207–240. Dr. W. Junk, The Hague.

- Bridel, S. E. 1812: Muscologia recentiorum supplementum 2. – Carolum G. Ettingerum, Gothae.
- Carine, M. A., Francisco-Ortega, J., Santos-Guerra, A. & Russell, S. J. 2004: Relationships of island and continental floras: molecular evidence for multiple colonisations into Macaronesia and subsequent back-colonisation of the continent in *Convolvulus* L. — *American Journal of Botany* 91: 1070–1085.
- De Sloover, J.-L. 1977: Note de bryologie africaine VIII. Neckera, Neckeropsis. — Bulletin du Jardin Botanique National de Belgique 47: 31–48.
- Désamoré, A., Laenen, B., Devos, N., Popp, M., González-Mancebo, J. M., Carine, M. & Vanderpoorten, A. 2011: Out of Africa: north-westwards Pleistocene expansions of the heather *Erica arborea*. — *Journal of Biogeography* 38: 164–176.
- Douzery, E. J. P., Pridgeon, A. M., Kores, P., Linder, H. P., Kurzweil, H. & Chase, M. W. 1999: Molecular phylogenetics of *Diseae* (Orchidaceae): a contribution from nuclear ribosomal ITS sequences. — *American Journal* of *Botany* 86: 887–899.
- Düll, R. 1992: Distribution of the European and Macaronesian mosses (Bryophytina). Annotations and progress. — Bryologische Beiträge 8/9: 1–223.
- Engler, A. 1879: Versuch einer Entwickllungsgeschichte, insbesondereder Florengebiete seit der Tertiärperiode. I. Die extratropischen Gebiete der nördlichen Hemisphäre. – W. Engelmann, Leipzig.
- Frey, W., Frahm, J. P., Fischer, E. & Lobin, W. 2006: *The liverworts, mosses and ferns of Europe.* Harley Books, Cochester.
- Goloboff, P., Farris, J. & Nixon, K. 2003: T.N.T.: Tree analysis using new technology. — Program and documentation available from the authors, and at www.zmuc.dk/ public/phylogeny/TNT/.
- González-Mancebo, J. M., Romaguera, F., Ros, R. M., Patiño, J. & Werner, O. 2008: Bryophyte flora of the Canary Islands: an updated compilation of the species list with an analysis of distribution patterns in the context of the Macaronesian Region. — *Cryptogamie*, *Bryologie* 29: 315–357.
- Guerra, J., Cano, M. J., Pérez Latorre, A. V., Ros, R. M. & Cabezudo, B. 2003: Flora brio-pteridofítica de los bosques lauroides de *Rhododendron ponticum* L. del Parque Natural de los Alcornocales (Cádiz-Málaga, España). — *Acta Botánica Malacitana* 28: 19–36.
- Guerra, J., Gil, J. A. & Varo, J. 1980: Dos briófitos nuevos para Europa continental. — *Boletim da Sociedade Broteriana* 54: 173–179.
- Guerra, J., Jiménez-Martínez, J. F. & Jiménez J. A. 2010: *Neckera baetica* sp. nov. (Neckeraceae, Bryophyta) from Southern Spain, based on morphological and molecular data. — *Nova Hedwigia* 91: 255–263.
- Hedenäs, L. 1992: Flora of Madeiran pleurocarpous mosses (Isobryales, Hypnobryalse, Hookeriales). — *Bryophytorum Bibliotheca* 44: 1–165.
- Hewitt, G. M. 2000: The genetic legacy of the ice ages. *Nature* 405: 907–913.
- Hill, M. O., Bell, N., Bruggeman-Nannenga, M. A., Brugués, M., Cano, M. J., Enroth, J., Flatberg, K. I., Frahm, J.-P.,

Gallego, M. T., Garilleti, R., Guerra, J., Hedenäs, L., Holyoak, D. T., Hyvönen, J., Ignatov, M. S., Lara, F., Mazimpaka, V., Muñoz, J. & Söderström, L. 2006: An annotated checklist of the mosses of Europe and Macaronesia. — *Journal of Bryology* 28: 198–267.

- Huelsenbeck, J. P. & Ronquist, F. 2001: MrBayes: Bayesian inference of phylogeny. – *Bioinformatics* 17: 754–755.
- Ignatov, M. S. & O. M. Afonina 1992: Checklist of mosses of the former USSR. – Arctoa 1: 1–85.
- Ignatov, M. S., Gardiner, A. A., Bobrova, V. K., Milyutina, I. A., Huttunen, S. & Troitsky, A. V. 2007: On the relationships of mosses of the order Hypnales, with special reference to taxa traditionally classified in the Leskeaceae. — In: Newton, A. E. & Tangney, R. (eds.), *Pleurocarpous mosses: systematics and evolution*: 177–214. Taylor & Francis CRC Press, Boca Raton.
- Juan, C., Emerson, B. C., Oromí, P. & Hewitt, G. M. 2000: Colonization and diversification: towards a phylogeographic synthesis for the Canary Islands. — *Trends in Ecology and Evolution* 15: 104–109.
- Kelchner, S. A. 2000: The evolution of non-coding chloroplast DNA and its application in plant systematics. — *Annals of the Missouri Botanical Garden* 87: 482–498.
- Mejías, J. A., Arroyo, J. & Marañón, T. 2007: Ecology and biogeography of plant communities associated with the post Plio-Pleistoicene relict *Rhododendron ponticum* subsp. *baeticum* in southern Spain. — *Journal of Biogeography* 34: 456–472.
- Müller, K. 2004: SeqState primer design and sequence statistics for phylogenetic DNA data sets. — Applied Bioinformatics 4: 65–69.
- Müller, K., Müller, J., Neinhuis, C. & Quandt, D. 2006: *PhyDE – Phylogenetic Data Editor*, v0.995. – Available at http://www.phyde.de.
- Olsson, S., Enroth, J., Buchbender, V., Hedenäs, L., Huttunen, S. & Quandt, D. 2011: *Neckera* and *Thamnobryum* (Neckeraceae, Bryopsida): Paraphyletic assemblages. – *Taxon* 60: 36–50.
- Patiño Llorente, J. & González-Mancebo, J. M. 2005: División Bryophyta. – In: Arechavaleta, M., Zurita N., Marrero, M. C. & Martín, J. L. (eds.), Lista preliminar de especies silvestres de Cabo Verde (hongos, plantas y animales terrestres): 34–37. Consejería de Medio Ambiente y Ordenación Territorial, Gobierno de Canarias, La Laguna.
- Posada, D. 2008: jModelTest: phylogenetic model averaging. — Molecular Biology and Evolution 25: 1253–1256.
- Rodríguez-Sánchez, F. & Arroyo, J. 2008: Reconstructing the demise of Tethyan plants: climate-driven range dynamics of *Laurus* since the Pliocene. — *Global Ecology and Biogeography* 17: 685–695.
- Rodríguez-Sánchez, F., Pérez-Barrales, R., Ojeda, F., Vargas, P. & Arroyo, J. 2008: The Strait of Gibraltar as a melting pot for plant biodiversity. — *Quaternary Science Reviews* 27: 2100–2117.

- Rodríguez-Sánchez, F., Guzmán, B., Valido, A., Vargas, P. & Arroyo, J. 2009: Late Neogene history of the laurel tree (*Laurus* L., Lauraceae) based on phylogeographical analyses of Mediterranean and Macaronesian populations. – *Journal of Biogeography* 36: 1270–1281.
- Ronquist, F. & Huelsenbeck, J. P. 2003: MrBayes 3: Bayesian phylogenetic inference under mixed models. – *Bioinformatics* 19: 1572–1574.
- Sérgio, C., Brugués, M., Cros, R. M., Casas, C. & Garcia, C. 2006: The 2006 Red List and an updated checklist of bryophytes of the Iberian Peninsula (Portugal, Spain and Andorra). – *Lindbergia* 31: 109–125.
- Simmons, M. P. & Ochoterena, H. 2000: Gaps as characters in sequence-based phylogenetic analyses. — *Systematic Biology* 49: 369–381.
- Sotiaux, A., Enroth, J., Olsson, S., Quandt, D. & Vanderpoorten, A. 2009: When morphology and molecules tell us different stories: a case-in-point with *Leptodon corsicus*, a new and unique endemic moss species from Corsica. – *Journal of Bryology* 31: 186–196.
- Stech, M. & Frahm, J.-P. 1999: The status of *Platyhypnidium mutatum* Ochyra & Vanderpoorten and the systematic value of the Donrichardsiaceae based on molecular data. — *Journal of Bryology* 21: 191–195.
- Sunding, P. 1979: Origins of the Macaronesian flora. In: Bramwell, D. (ed.), *Plants and islands*: 13–40. Academic Press, London.
- Taberlet, P., Gielly, L., Pautou, G. & Bouvet, J. 1991: Universal primers for amplification of three non-coding regions of chloroplast DNA. — *Plant Molecular Biology* 17: 1105–1109.
- Thompson, J. D. 2005: Plant evolution in the Mediterranean. — Oxford University Press, Oxford.
- Tsubota, H., Arikawa, T., Akiyama, H., De Luna, E., Gonzales, D., Higuchi, M. & Deguchi, H. 2002: Molecular phylogeny of hypnobryalean mosses as inferred from a large-scale dataset of chloroplast *rbcL*, with special reference to the Hypnaceae and possibly related families. — *Hikobia* 13: 645–665.
- Van der Wijk, R., Margadant, W. D. & Florchütz, P. A. 1964: Index muscorum 3. Hypnum-O. – *Regnum Vegetabile* 33: 1–529.
- Vanderpoorten, A., Laenen, B., Rumsey, F., Gonzalez-Mancebo, J. M. & Gabriel, R. 2011: Dispersal, diversity and evolution of the Macaronesian cryptogamic floras. — In: Bramwell, D. (ed.), *Plants and islands*, 2nd ed. Cambridge University Press, Cambridge. [In press].
- Vogel, J. C., Rumsey, F. J., Schneller, J. J., Barrett, J. A. & Gibby, M. 1999: Where are the glacial refugia in Europe? Evidence from pteridophytes. — *Biological Journal of the Linnean Society* 66: 23–37.
- Werner, O., Ros, R. M. & Guerra, J. 2002: Direct amplification and NaOH extraction: two rapid and simple methods for preparing bryophyte DNA for polymerase chain reaction (PCR). – *Journal of Bryology* 24: 127–131.

Appendix. Herbarium vouchers, studied specimens' geographic origin and Genbank accession numbers (ITS; *trn*L-F).

Alleniella besseri. **1**. S-B106203, Sweden, Värmland (JF690782; JF690819); **2**. S-B125094, Sweden, Dalarna (JF690783; JF690820).

- Alleniella complanata. 1. MUB-30336/TFCBry-15241, Spain, Canary Islands, La Gomera (JF690784; JF690821); 2. MUB-30337/TFCBry-15233, Spain, Canary Islands, La Gomera (JF690785; JF690822); 3. MUB-30335/TFCBry-15256, Spain, Canary Islands, Tenerife (JF690786; JF690823); 4. MUB-30334/TFCBry-15253, Spain, Canary Islands, Tenerife (JF690787; JF690824); 5. MUB-34552, Spain, Huesca (JF690788; JF690825); 6. S-B9726, Portugal, Madeira (JF690789; JF690826).
- Exsertotheca baetica. 1. MUB-34553, Spain, Cádiz (JF690800; JF690841); 2. MUB-34554, Spain, Cádiz (-; JF690842);
 3. Autonomous University of Madrid Bryophyte Herbarium, Spain, Cádiz (-; JF690843); 4. Autonomous University of Madrid Bryophyte Herbarium, Spain, Cádiz (-; JF690844); 5. S-B100277, Spain, Cádiz (-; JF690845).
- Exsertotheca crispa. 1. Autonomous University of Madrid Bryophyte Herbarium, Spain, Cuenca (JF690796; JF690834); 2.
 S-B125096, Germany, Baden-Württemberg (JF690795; JF690833); 3. MUB-10055, Spain, Cádiz (JF690797; JF690835);
 4. S-B44528, Portugal, Beira Litoral, Coimbra (JF690798; JF690836); 5. S-B111464, Switzerland, Tessin (JF690799; JF690837);
 6. MUB-34555, Spain, Cádiz (-; JF690838);
 7. Autonomous University of Madrid Bryophyte Herbarium, Spain, Cádiz (-; JF690839);
 8. Autonomous University of Madrid Bryophyte Herbarium, Spain, Guadalajara (-; JF690840).
- Exsertotheca intermedia. 1. S-B9730, Portugal, Madeira (JF690816; JF690861); 2. MUB-310007/TFCBry-17120, Spain, Canary Islands, Fuerteventura (JF690801; JF690840); 3. MUB-310008/TFCBry-17123, Spain, Canary Islands, Fuerteventura (JF690802; JF690847); 4. MUB-310009/TFCBry-17126, Spain, Canary Islands, Fuerteventura (JF690802; JF690847); 4. MUB-310009/TFCBry-17126, Spain, Canary Islands, Fuerteventura (JF690803; JF690848);
 MUB-31001/TFCBry-15242, Spain, Canary Islands, La Gomera (JF690804; JF690849);
 MUB-31001/TFCBry-15242, Spain, Canary Islands, La Gomera (JF690805; JF690850);
 MUB-31006/TFCBry-15234, Spain, Canary Islands, La Gomera (JF690806; JF690851F);
 MUB-31005/TFCBry-15801, Spain, Canary Islands, La Gomera (JF690807; JF690852);
 MUB-31004/TFCBry-15250, Spain, Canary Islands, Tenerife (JF690808; JF690853);
 MUB-31004/TFCBry-15252, Spain, Canary Islands, Tenerife (JF690809; JF690854);
 S-B42781, Portugal, Azores, São Miguel (JF690810; JF690855);
 S-B42782, Portugal, Azores, São Miguel (JF690811; JF690856);
 AZU, Portugal, Azores, Santa María (JF690812; JF690857);
 MUB-30332/TFCBry-15264, Portugal, Madeira (JF690813; JF690858);
 E-00266405, Portugal, Madeira (JF690814; JF690859);
 MUB-30333/TFCBry-15263, Portugal, Madeira (JF690815; JF690859);

Leptodon smithii. MUB-30331/TFCBry-15246, Spain, Canary Islands, La Palma (JF690781; JF690818).

Neckera cephalonica. 1. S-B9806, Portugal, Madeira (JF690793; JF690831); 2. S-B9824, Portugal, Madeira (JF690794; JF690832).

Neckera menziesii. GenBank FM161167; FM210305.

- Neckera pennata. GenBank FM161169; AM990414.
- Neckera pumila. 1. Autonomous University of Madrid Bryophyte Herbarium, Spain, Málaga (JF690790; JF690827);
 2. S-B44531, Portugal, Beira Litoral, Coimbra (JF690791; JF690828);
 3. S-B9798, Portugal, Madeira (JF690792; JF690829). Neckera cf. pumila. Autonomous University of Madrid Bryophyte Herbarium, Spain, Cádiz (-; JF690830).